

National Institute of Food and Agriculture

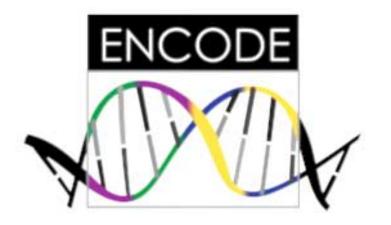
# Functional Annotation of the Animal Genome FAANG

American Society of Animal Science, Canadian Society of Animal Science
Midwestern Section
March 11, 2019









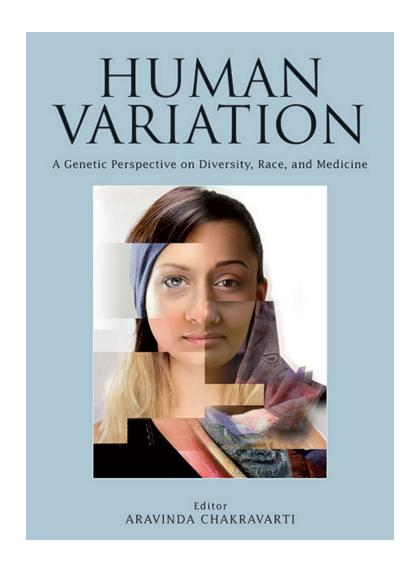
# **ENCODE:** Encyclopedia of DNA Elements



# **Biological Variation**

• What are the sources of variation in biological traits?







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- Caused by
  - Genetics
  - Environment

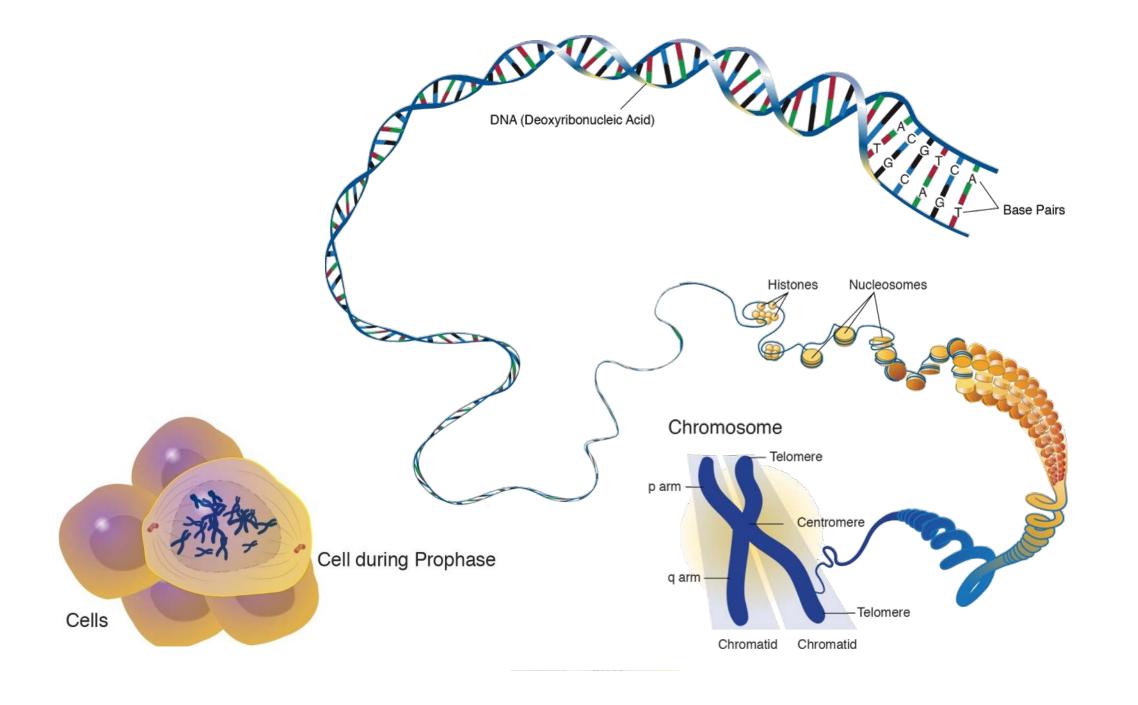


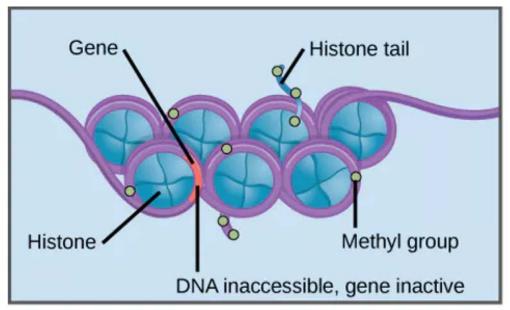
# **Biological Variation**

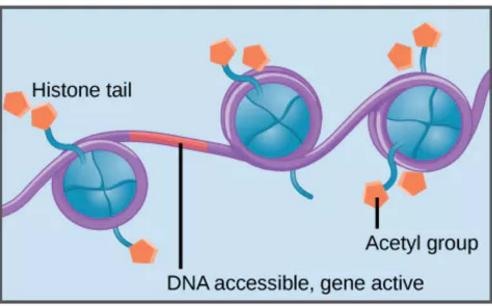
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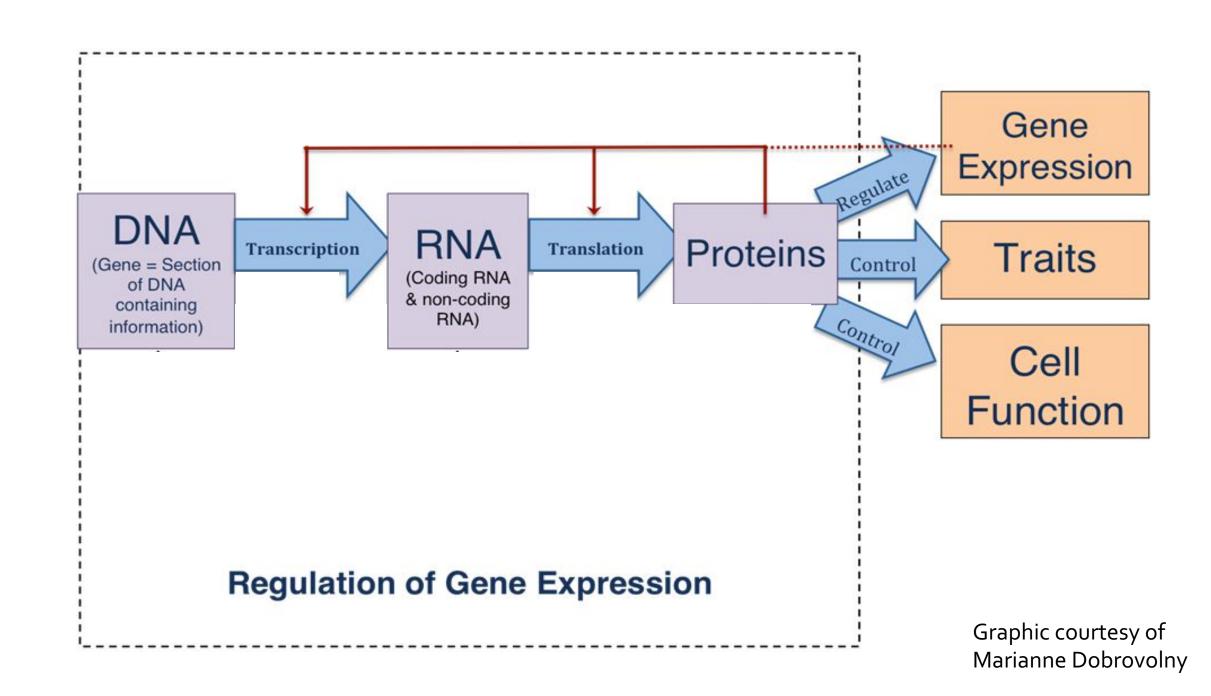








- Methylation histone & DNA causes nucleosome to be tightly pack
- Transcription factors cannot bind DNA & genes are NOT expressed
- Histone acetylation results in loose packaging
- Transcription factor CAN bind DNA & genes can be expressed





National Institute of Food and **Aariculture** 

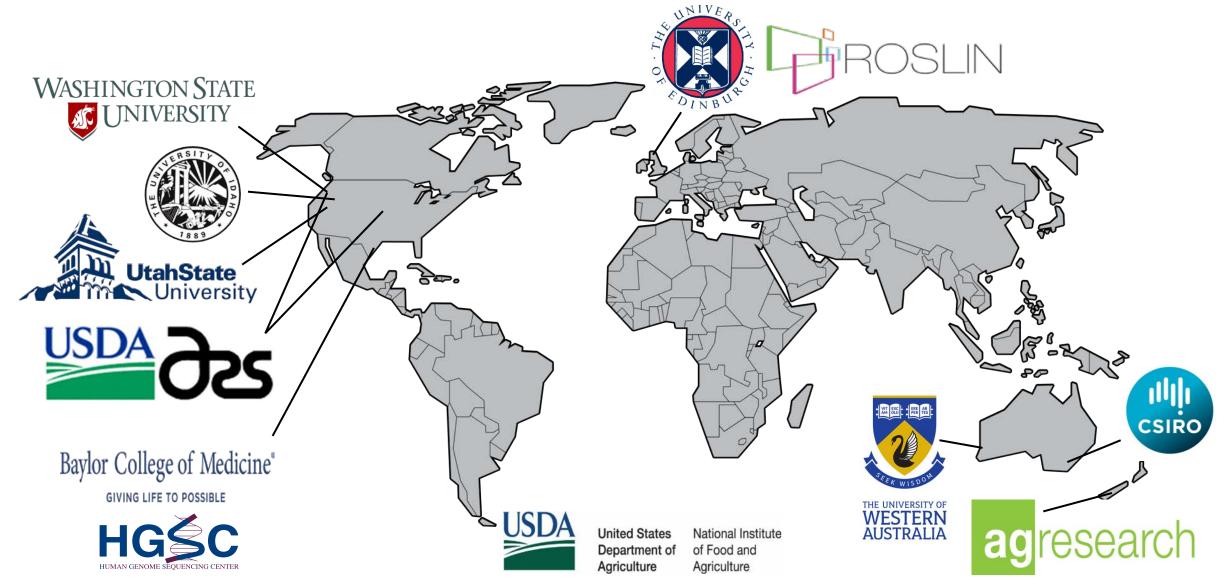
United States



Brenda Murdoch (University of Idaho), Stephen White (USDA, ARS, Washington State University), Michelle Mousel (USDA, ARS ADRC), Alisha Massa (Washington State University), Kim Worley (Baylor College of Medicine), Alan Archibald (Roslin Institute), Emily Clark (Roslin Institute), Brian Dalrymple (University of Western Australia), James Kijas (CSIRO), Shannon Clarke (AgResearch), Rudiger Brauning (AgResearch), Timothy Smith (USDA, ARS MARC), Tracey Hadfield (Utah State University), Noelle Cockett (Utah State University).

# Acknowledgements





# Objectives

1. Develop a deep and robust dataset of **transcribed** elements in the sheep genome.

2. Develop a deep and robust dataset of transcription regulatory features in the sheep genome.

3. Annotation of the ovine genome reference assembly.

4. Provide ovine FAANG data and support to the public.

# Materials and Methods

- We generated a new high quality reference genome sequence de novo assembled using (PacBio) sequence and Hi-C data.
  - The 2.87 Gb assembly (Oar\_rambouillet\_v1.0, GCA\_002742125.1)
- 100 tissues collected (snap & slow frozen) from Benz 2616 in April, 2016 at USU
  - BioProject PRJNA414087
- FAANG Annotations from the same animal - Oar\_rambouillet\_v1.0

Baylor College of Medicine®

GIVING LIFE TO POSSIBLE

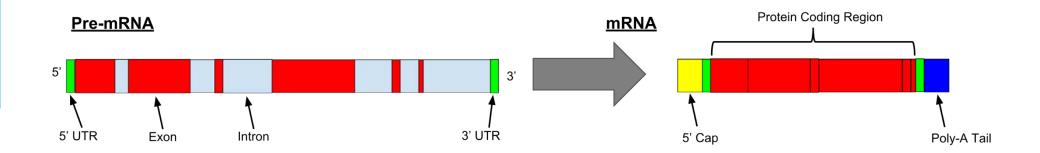




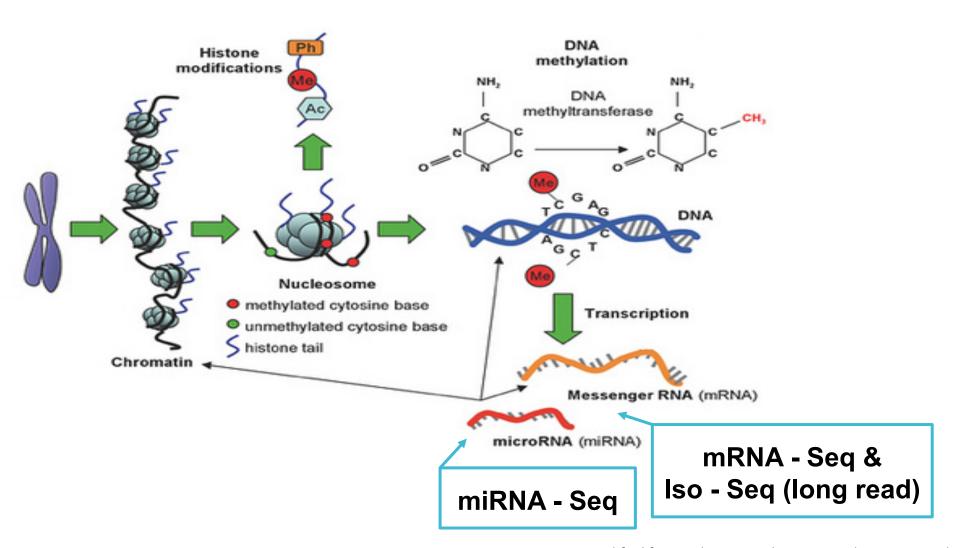
Rambouillet ewes Benz 2616 (right)

# Objectives

- 1. Develop a deep and robust dataset of **transcribed** elements in the sheep genome.
  - Three types of transcriptome data on tissues
    - 1) Using poly-A+ mRNAs transcript sequence
    - 2) Small noncoding microRNAs sequence
    - 3) Full transcript (ISO) sequence PacBio



# Sheep FAANG Assays



# Objectives

- 1. Develop a deep and robust dataset of **transcribed** elements in the sheep genome.
  - RNA sequencing and transcriptome analyses is in progress.

Transcript	Tissues run by Kim Worley	Tissues run by Tim Smith	Total Tissues
mRNA	29	31	60
miRNA	30	-	30
Iso-Seq	5	10	15

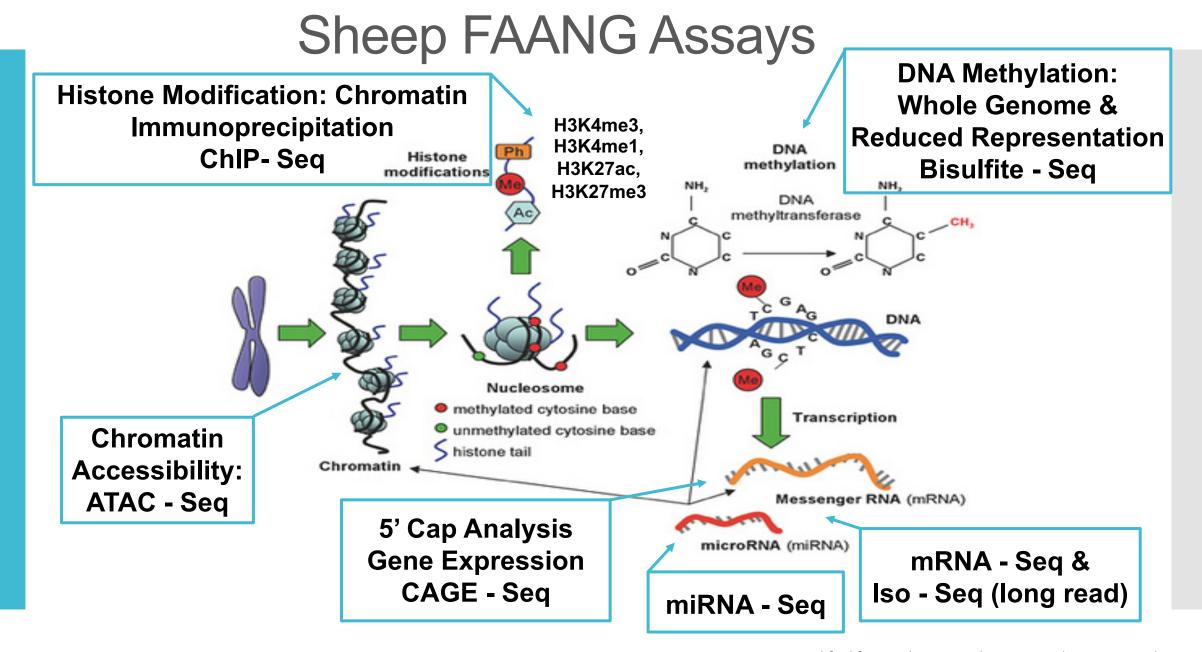




# Objectives

1. Develop a deep and robust dataset of **transcribed** elements in the sheep genome.

- 2. Develop a deep and robust dataset of transcription regulatory features in the sheep genome.
  - 1) Cap Analysis of Gene Expression: CAGE Sequencing
  - 2) Chromatin accessibility: ATAC Sequencing
  - 3) Histone modification: Chromatin Immunoprecipitation Sequencing
  - 4) DNA methylation: Reduced Representation Bisulfite and Whole Genome Bisulfite Sequencing



# Regulatory Feature Assays

- CAGE assay & analyses, lead by Emily Clark and Alan Archibald is currently in progress.
  - In total 58 samples of mRNA were shipped to Scotland
  - CAGE assay sequenced on the HiSeq 2500 platform
  - To date 40 different tissues have been analyzed

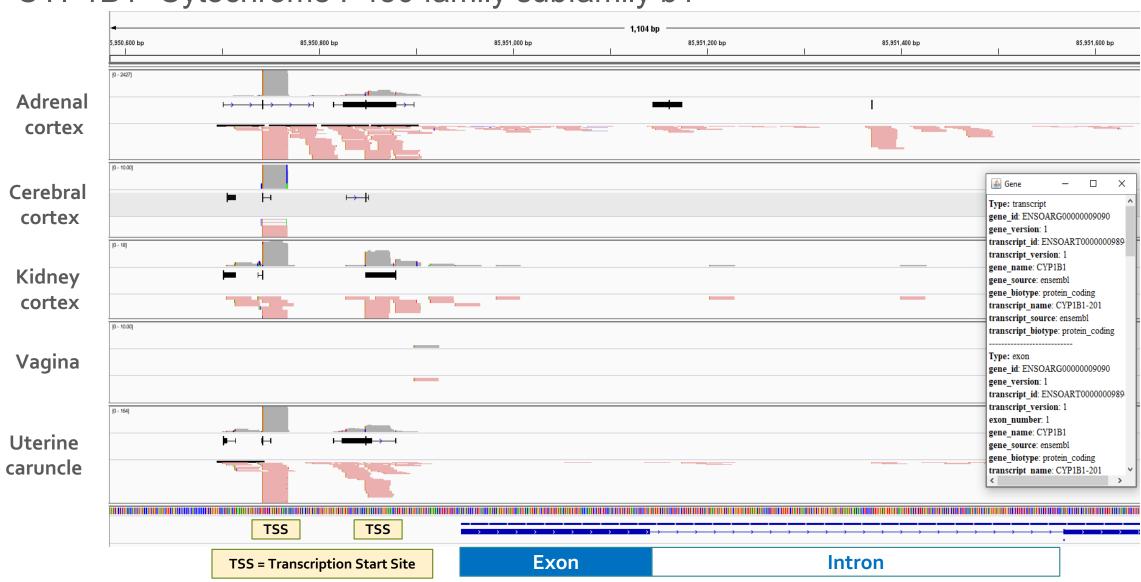






# CAGE – Sequence

CYP1B1- Cytochrome P450 family subfamily b1



# Regulatory Feature Assays

- ChIP-Seq & ATAC-Seq, are in progress in collaboration with Brenda Murdoch, Stephen White and Michelle Mousel.
- Separate pilot projects are underway to ensure protocols are working for sheep tissues.
  - Initial pilot experiment to develop protocols are being analyzed
  - 3 tissues from 2 female and 2 males

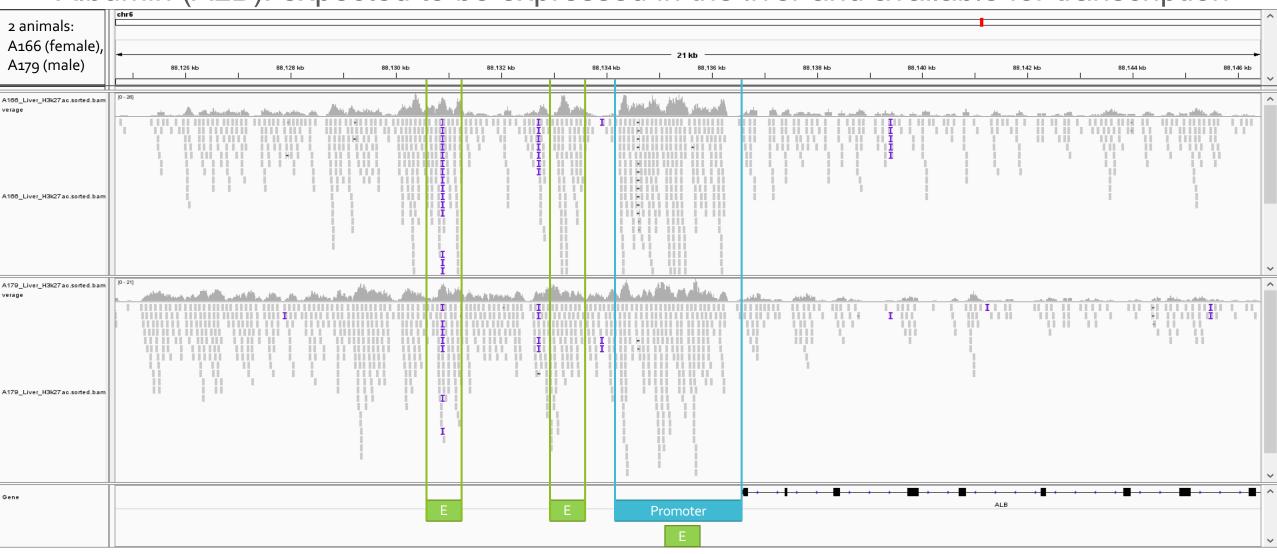






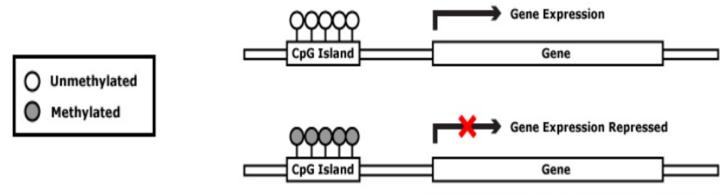
# ChIP – Sequence for H3K27ac

Albumin (ALB): expected to be expressed in the liver and available for transcription



# Regulatory Feature Assays

- DNA methylation, is being lead by Shannon Clarke and Rudiger Brauning.
  - Reduced Representation Bisulfite Sequencing
    - on the same 60 tissues as the RNA sequence
  - Whole Genome Bisulfite Sequencing
    - for 22 core FAANG tissues





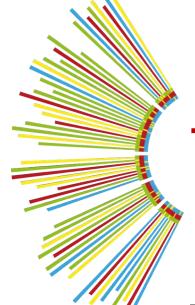
# Summary

1. Transcriptome data for 60 tissues.

- 2. Transcription regulatory features in the sheep genome.
  - CAGE for 40 tissues completed
- 3. Annotation of the ovine genome reference assembly.

4. Provide ovine FAANG data and support to the public.







Pablo Ross <sup>1</sup>, Stephanie McKay <sup>2</sup>, Clare Gill <sup>3</sup>, Monique Rijnkels <sup>3</sup>, Brenda Murdoch <sup>4</sup>, Tim Smith <sup>5</sup>, Huaijun Zhou <sup>1</sup>, Zhihua Jiang <sup>6</sup>, James Reecy <sup>7</sup>, Wansheng Liu <sup>8</sup>, Honglin Jiang <sup>9</sup>, Milton Thomas <sup>10</sup>

<sup>1</sup> University of California Davis, <sup>2</sup> University of Vermont, <sup>3</sup>Texas A&M University, <sup>4</sup> University of Idaho, <sup>5</sup> USDA-ARS, <sup>6</sup> Washington State University, <sup>7</sup> Iowa State University, <sup>8</sup> Pennsylvania State University, <sup>9</sup> Virginia Tech, <sup>10</sup> Colorado State University



### **BIOLOGICAL SAMPLES**



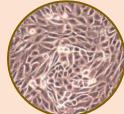
L1 adult tissues (UC Davis)

Holstein mammary tissue (Texas A&M)

L1 fetal tissues (U of Idaho)



Cultured cells & embryos (UC Davis)



Cultured cells (Virginia Tech)

### **OBJECTIVE I**

**Transcriptome Annotation** 

RNA-seq Small RNA-seq RAMPAGE ATTS-seq

### **OBJECTIVE II**

**Chromatin State Annotation** 

ATAC-seq ChIP-seq WGBS

### **OBJECTIVE III**

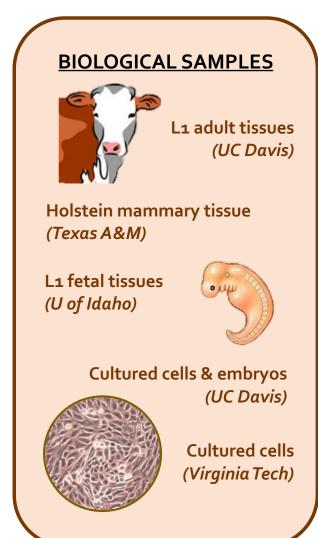
Data Integration
Functional Element Annotation
Public Distribution

### **OBJECTIVE IV**

Integrate Functional Maps with GWAS analysis

Using two populations:

- 1) Multigenerational GWAS For production efficiency traits
- 2) Growth and carcass data on >4,000 genotyped cattle
- Use functional annotation to narrow genomic regions of interest
- Identify genomic variants associated with production, growth, carcass traits
- Use this information to identify causal variants (beyond the scope of this project)



- 40 adult tissues from L1 Hereford line
- 5 stages of mammary gland development
- 8 fetal tissues from L1 Hereford line
- 8 primary cell lines

Expressed regions			
RNA-seq	transcripts - variants		
smRNA- seq	Small transcript		
RAMPAGE	Transcription start sites		
WTTS-seq	Transcription termination sites		
lso-seq	Large transcripts		

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**Transcriptome Annotation** 

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Chromatin State			
ATAC-seq	Open chromatin		
H <sub>3</sub> K <sub>4</sub> me <sub>3</sub>	Active promoters		
H <sub>3</sub> K <sub>2</sub> 7me <sub>3</sub>	Repression		
H <sub>3</sub> K <sub>4</sub> me <sub>1</sub>	Active enhancers		
Hakazac	Enhancers &		
пзк2/ас	promoters		
CTCF	Insulators &		
CICI	promoters		
H <sub>3</sub> K <sub>9</sub> me <sub>3</sub>	Heterochromatin		
H <sub>3</sub> K <sub>3</sub> 6me <sub>3</sub>	Active gene bodies		
WGBS	DNA methylation		
	H3K4me3 H3K27me3 H3K4me1 H3K27ac  CTCF H3K9me3 H3K36me3		

### **BIOLOGICAL SAMPLES**



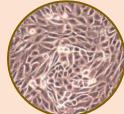
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# Summary

- Transcriptome for 40 adult tissues from L1 Hereford line is close to completion
- Fetal tissues (3 male, 1 female) from L1 Hereford line have been collected
- Assay for primary cell lines are underway

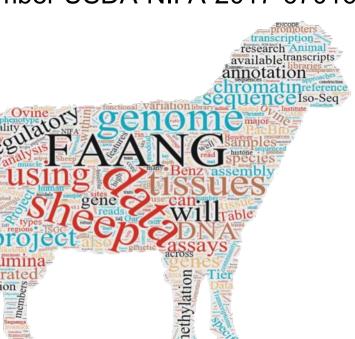
# Conclusions

- 1. Develop a deep and robust dataset of **transcribed** elements in the genome.
  - Sheep (60 tissues) & Cattle (40 tissues, mammary & fetal, cells)
- 2. Develop a deep and robust dataset of transcription **regulatory features** in the genome.
  - Sheep & Cattle
- 3. Annotation of the genome reference assembly.
- 4. Provide FAANG data and support to the public.

# Acknowledgements



This material is based upon work that is supported by the National Institute of Food and Agriculture, U.S. Department of Agriculture, award number USDA-NIFA-2017-67016-26301.







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