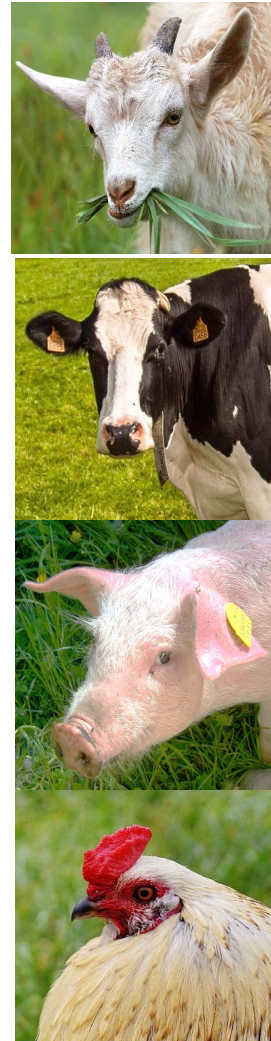


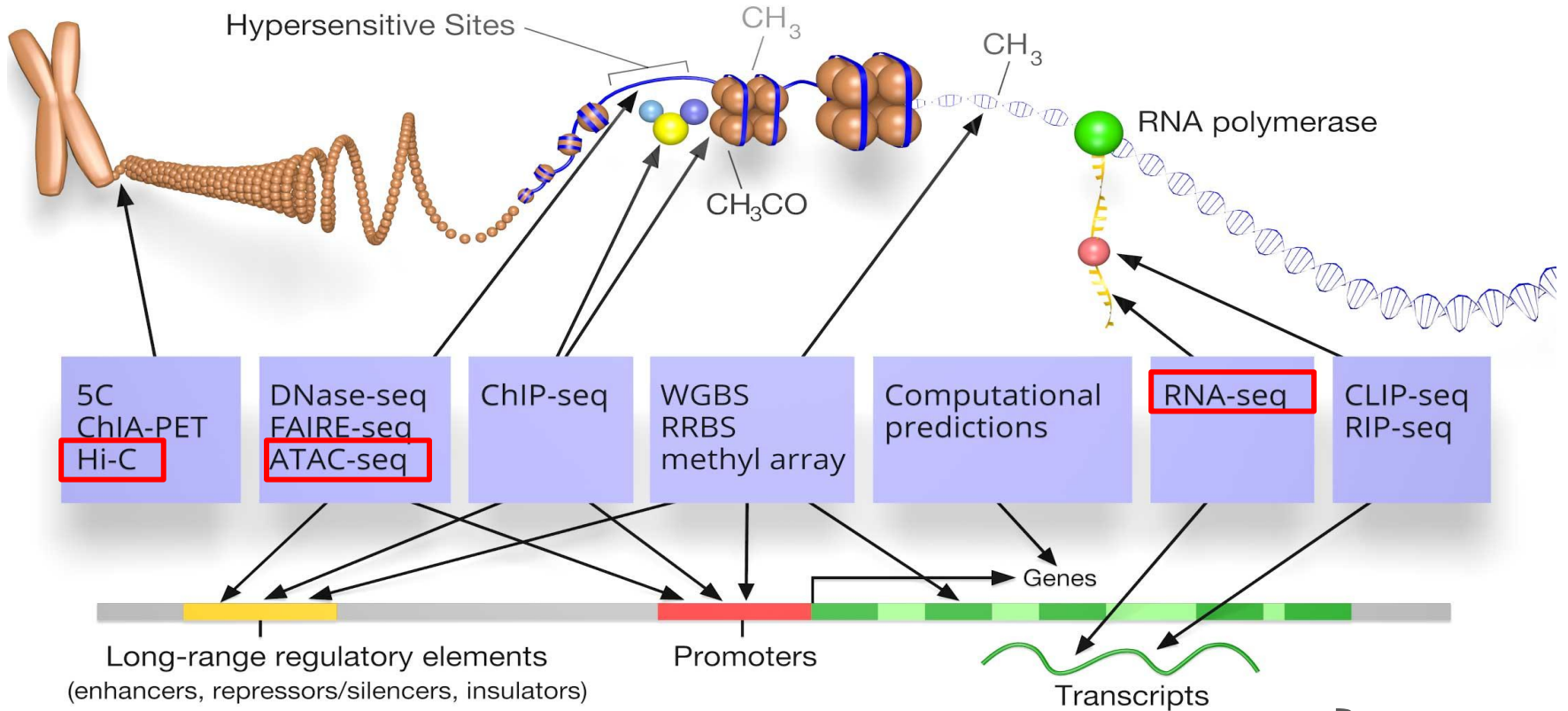
An update on the FAANG pilot project FR-AgENCODE

S. Foissac, S. Djebali, **Andrea Rau**, S. Lagarrigue,
H. Acloque, E. Giuffra, and the FR-AgENCODE group

INRA - {Jouy en Josas, Toulouse, Rennes} - France



FR-AgENCODE data



{2 ♂ + 2 ♀}
×
Liver
CD4
CD8

Based on an image by Darryl Leja (NHGRI), Ian Dunham (EBI), Michael Pazin (NHGRI)

Overview of FR-AgENCODE analyses

Multi-species single data analyses:

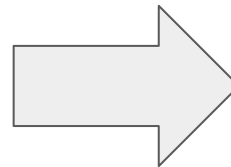
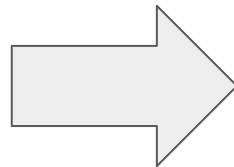
- Expression (**RNA-seq**)
- Open chromatin (**ATAC-seq**)
- 3D chromatin conformation (**Hi-C**)



} Quantification, normalization, differential analysis, clustering, ...

Multi-species integrative analyses:

- Expression + open chromatin + 3D chromatin conformation
- Expression + open chromatin

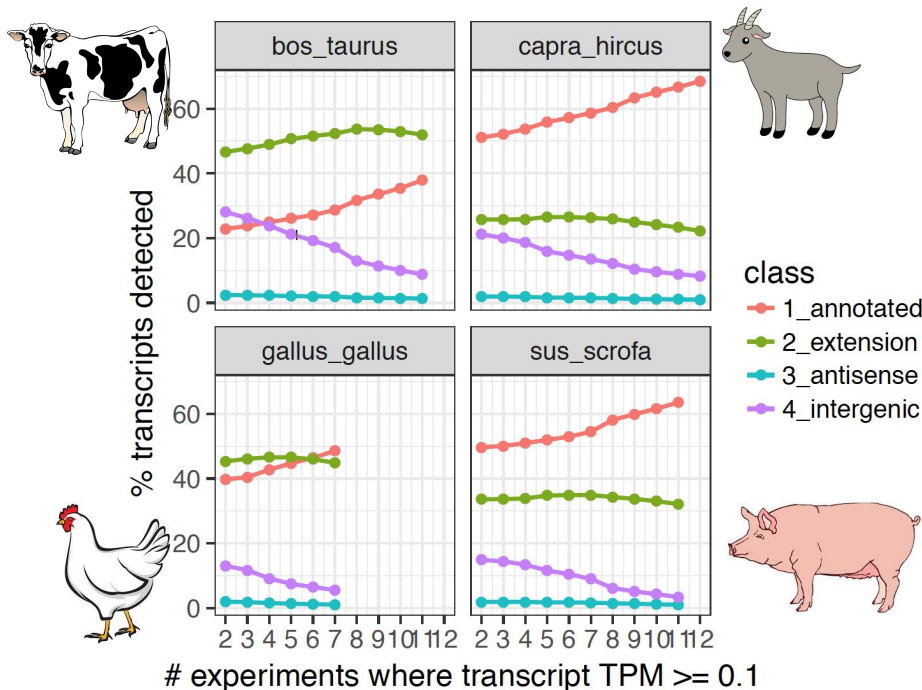


RNA-seq identifies many novel transcripts

Species	Genome / Gene annotation	Annotated Transcripts		Number of novel transcripts detected*	Number of novel lncRNAs**	
		Total number	Detected*			
			#			% of total
Bos taurus	UMD 3.1 / Ensembl 90	26,740	16,100	60.2	65,539	
Capra hircus	CHIR_ARS 1 / NCBI	53,266	34,442	64.7	38,197	
Gallus gallus	GalGal 5 / Ensembl 90	38,118	22,180	58.2	34,852	
Sus scrofa	SScrofa 11.1 / Ensembl 90	49,448	29,786	60.2	39,032	

* with TPM ≥ 0.1 in ≥ 2 samples / ** with at least 2 exons and classified by FEELnc

Directed, 2 x 150bp, 100 million read pairs / sample



Mapped reads:

- Exonic: 50-80%
- Intronic: 10-20%
- Intergenic: 20% for cattle/chicken, 10% for goat/pig

Differentially expressed genes reflect underlying biology

Two per-gene (TPM > 0.1 in at least 2 samples) differential models:

- **Tissue effect**, blocking on individuals:

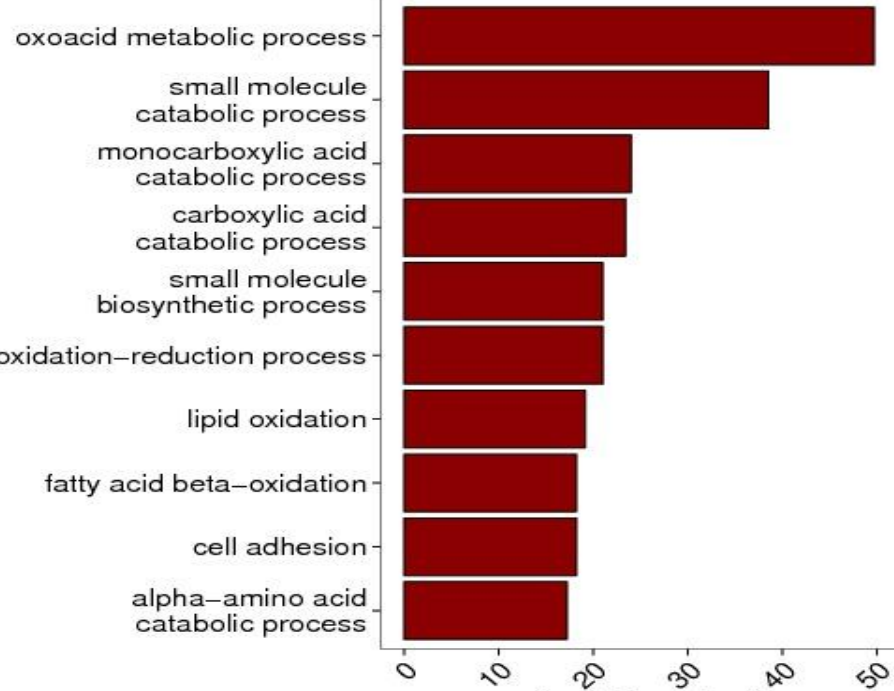
$$\text{expression} \sim \text{individual} + \text{tissue}$$

- **Tissue & tissue-specific sex effects**, blocking on (nested) individuals:

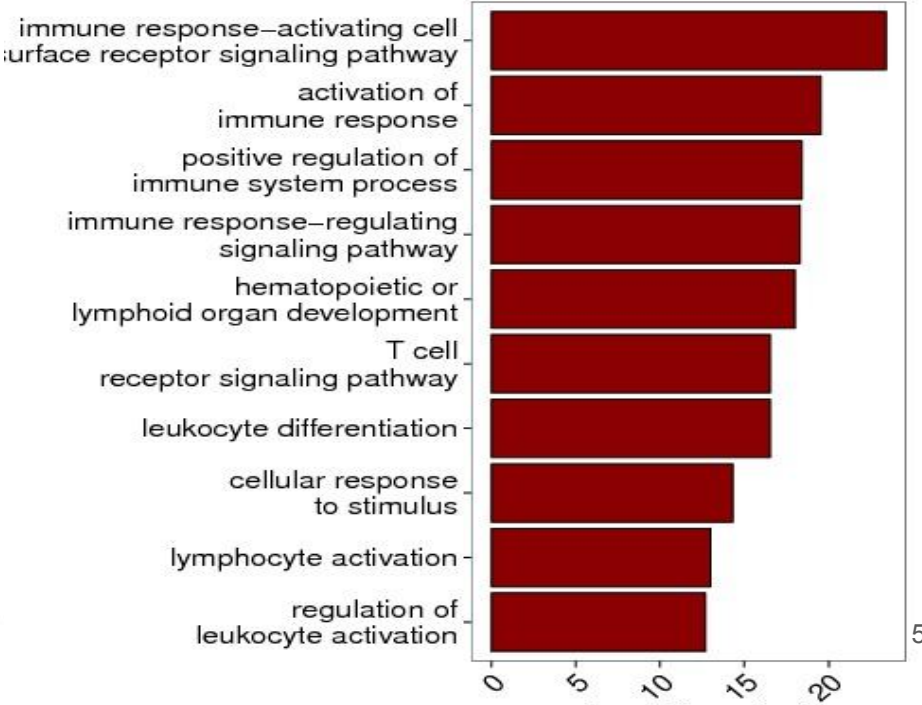
$$\text{expression} \sim \text{sex} + \text{tissue:sex} + \text{individual:sex}$$

- Generally large differences for liver vs CD4/CD8, smaller differences for CD4 vs CD8 or tissue-specific ♂ vs ♀
- GO (BP) enrichment for genes consistently DE across species:

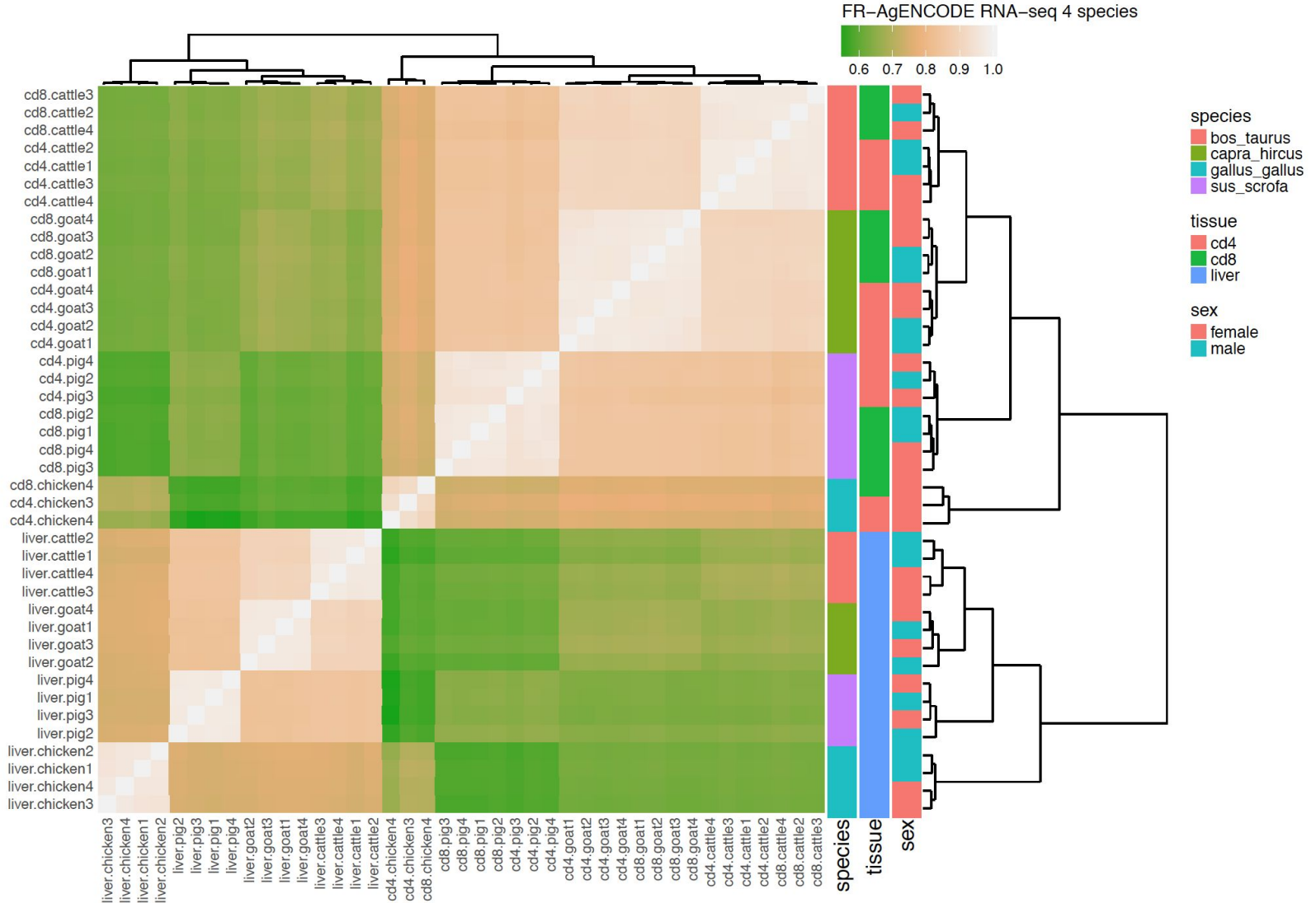
Over-expressed in **liver**



Over-expressed in **immune cells**

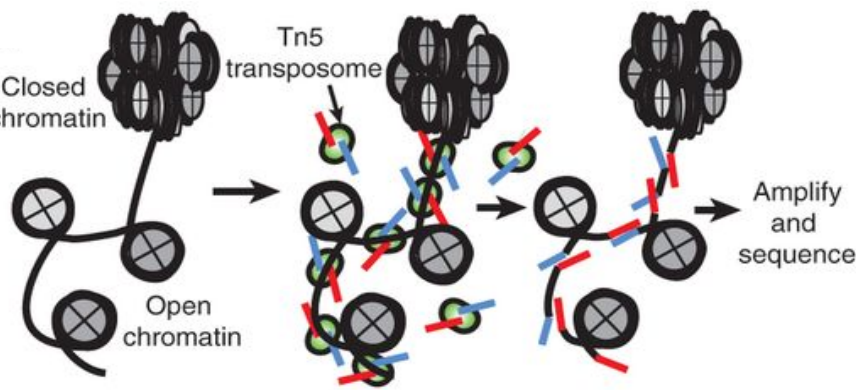


Cross-species & cross-tissue transcriptome clustering



- Liver clearly separated from CD4/CD8 cells, and species cluster within cell types (liver vs T-cells)

ATAC-seq identifies pertinent regions of open chromatin

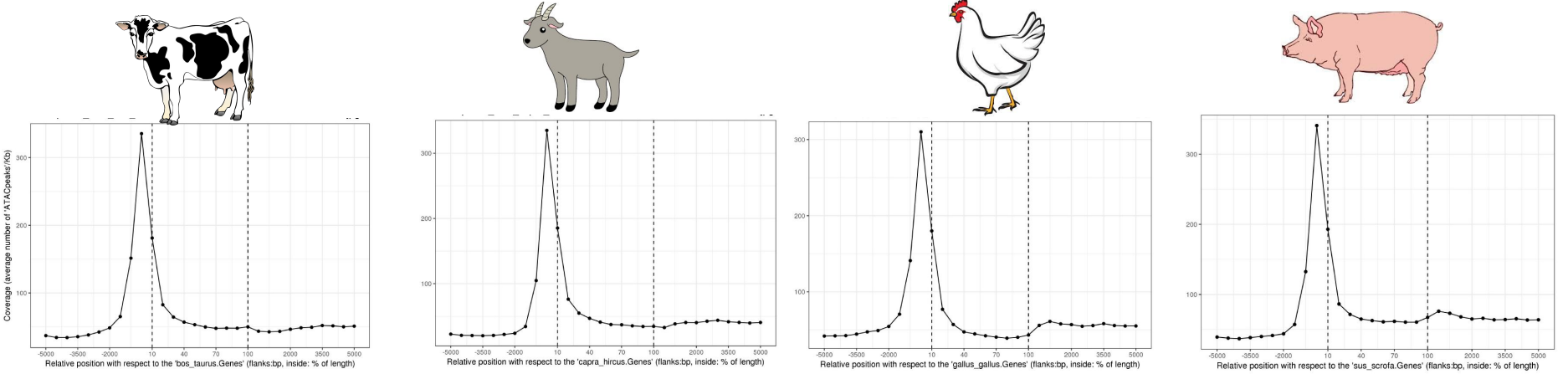


[Buenrostro et al., Nature Methods, 2013]

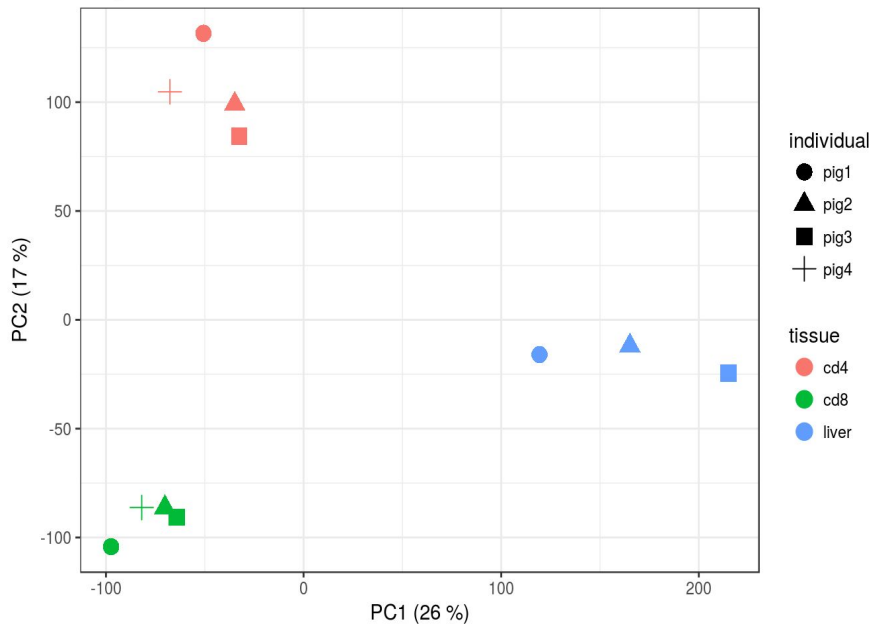
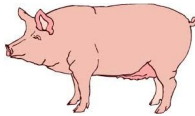
Species	Number of ATAC-seq peaks	Genome size (bp)	ATAC-seq peak coverage	
			# bp	% of genome
Bos taurus	104,986	2,670,422,299	80,562,624	3.02
Capra hircus	74,806	2,922,813,246	57,043,999	1.95
Gallus gallus	119,894	1,230,258,557	51,000,066	4.15
Sus scrofa	149,334	2,501,912,388	106,645,814	4.26

* 50 million read pairs / sample

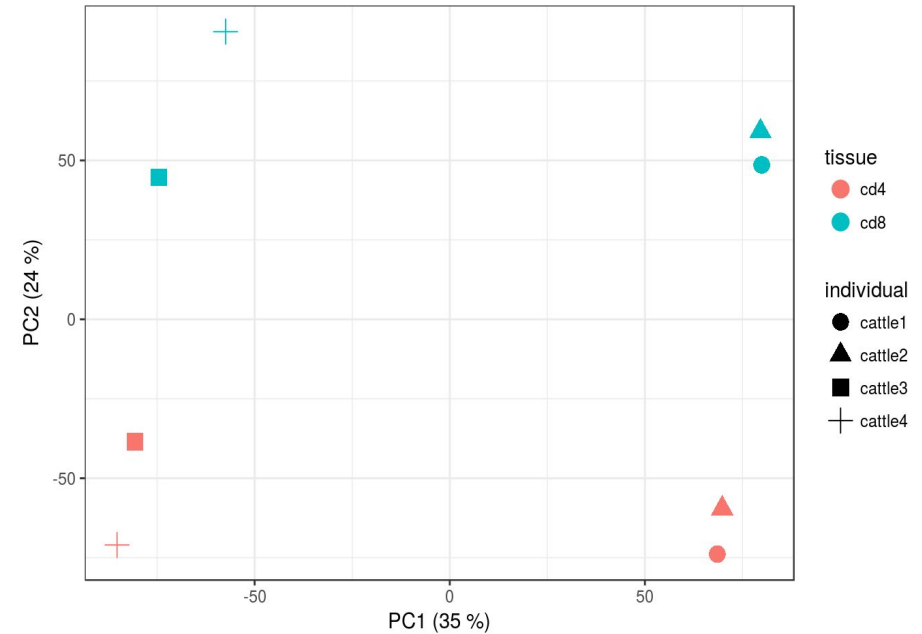
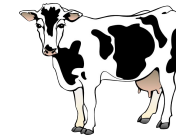
- 3 major peak classes: **promoter regions** (TSS +/- 1kb), **intronic**, **intergenic**
- Significant proportion of peaks in ATAC-seq reads found in promoter region (36-66%)
- For all species, peaks close to TSS (+/- 5kb) are most ubiquitous open chromatin



ATAC-seq sample clustering: Liver vs immune cells, ♂ vs ♀ in immune cells

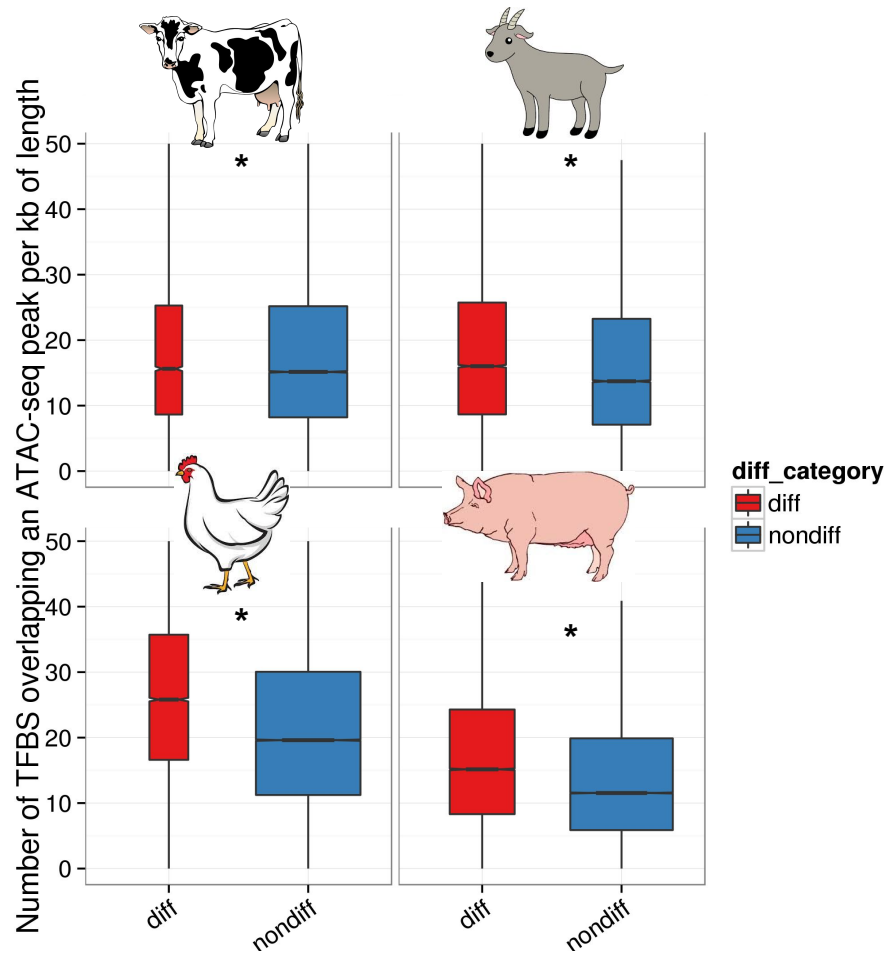


- PC1: liver versus CD4/CD8
- PC2 : CD4 vs CD8



- PC1: males vs females (note: absence of liver samples)
- PC2: CD4 vs CD8

Differential ATAC-seq peaks are more likely to be regulatory

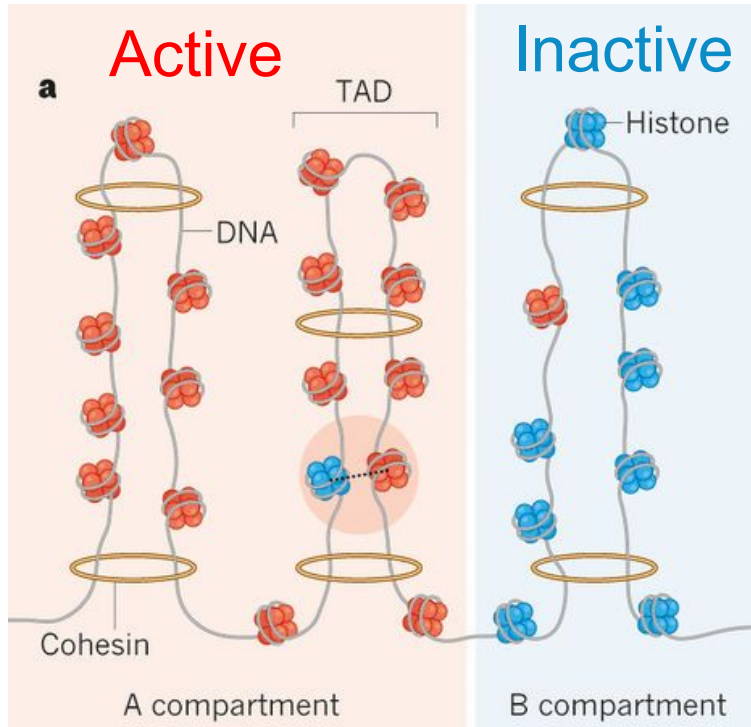


Between-tissue **differential** ATAC-seq peaks have a higher **TFBS density** than nondifferential peaks

(Wilcoxon test, p -value $< 10^{-15}$)

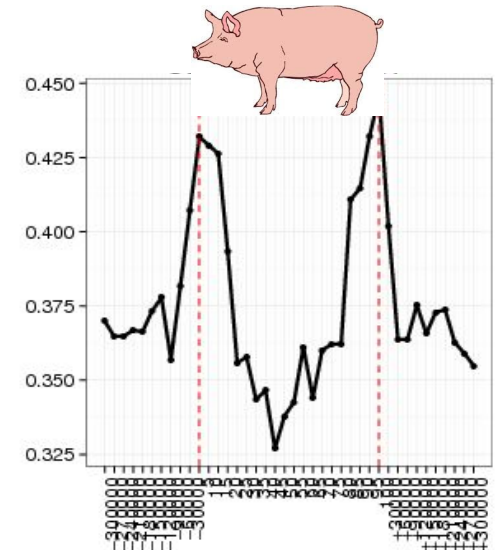
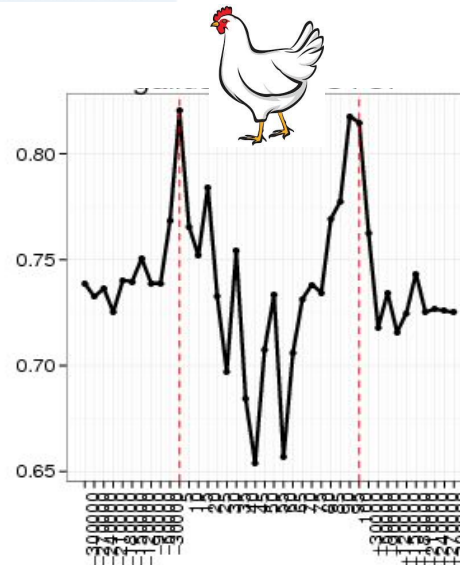
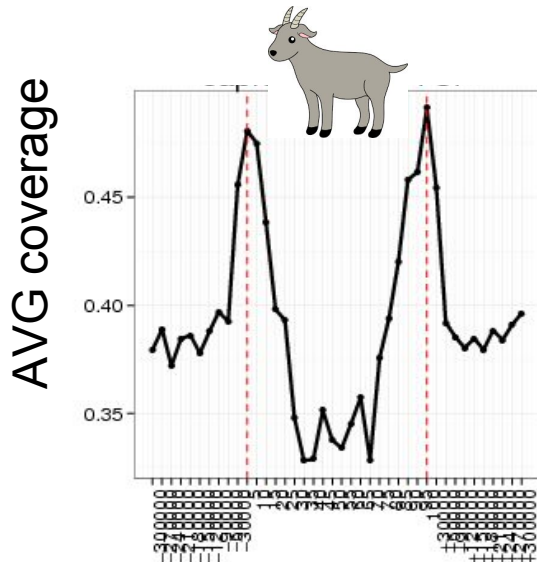
→ Differential ATAC-seq peaks are more likely to have a **regulatory** role

Hi-C for 3D genomic structures

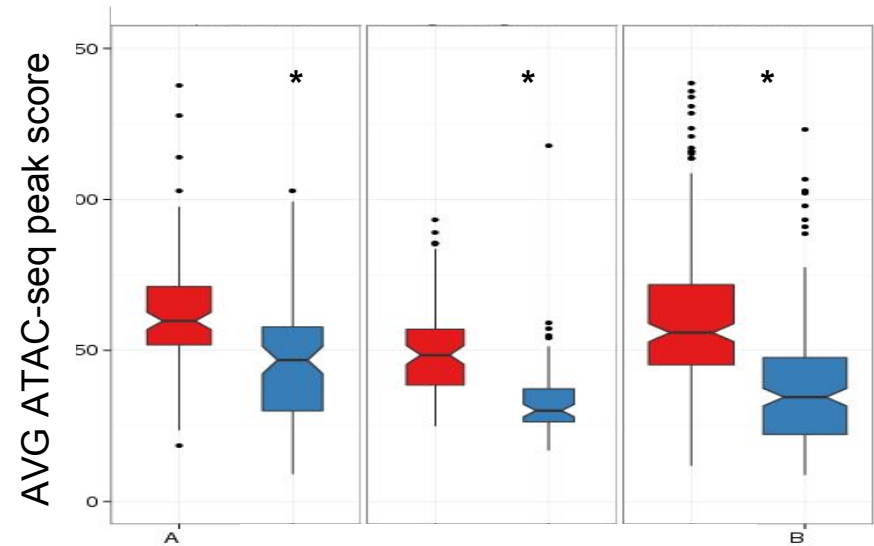
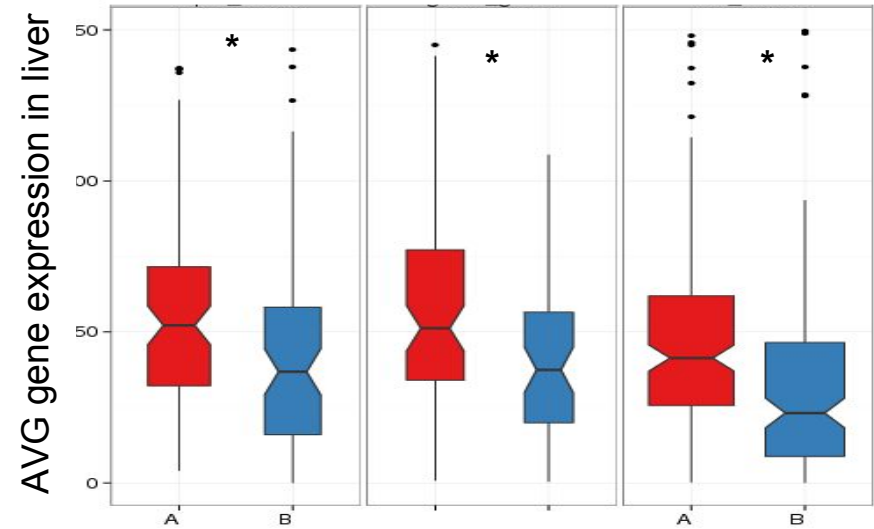
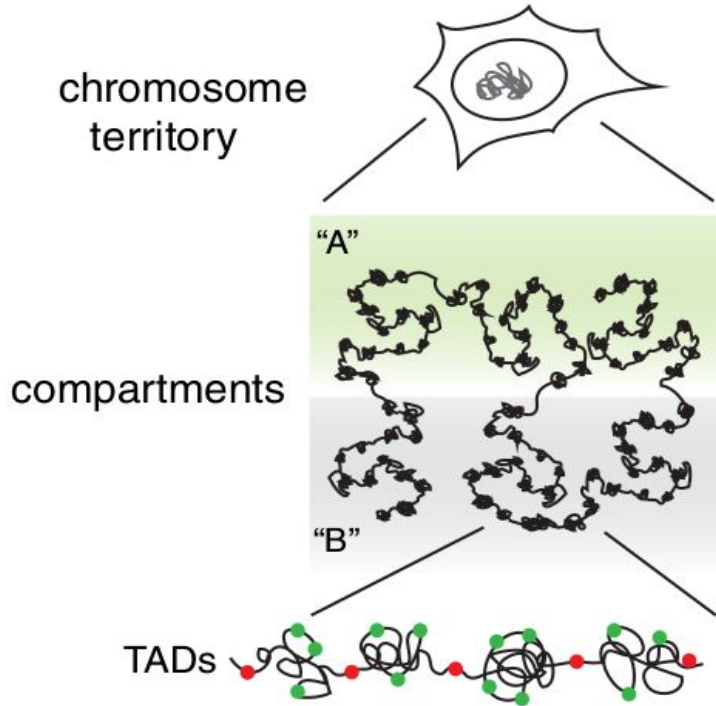
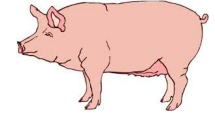


Several levels for 3D conformation:

- TADs (Topologically associating domains)
 - Compartments
 - Enhancer-promoter interactions
- Liver x 4 animals x 3 species (all but cattle)
 - 180 million read pairs / sample
 - Predicted CTCF binding sites peak at TAD boundaries ✓



RNA-seq + ATAC-seq + Hi-C consistency

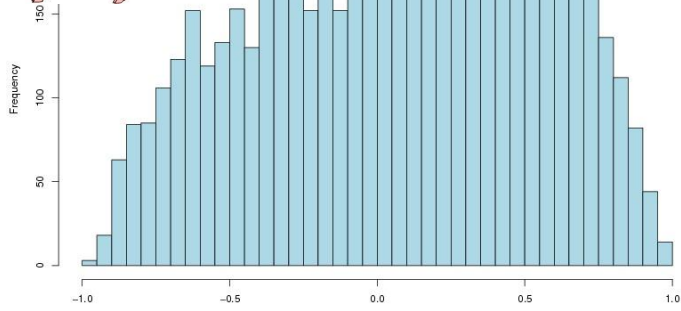
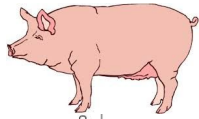


A compartments: open, expressed
B compartments: closed, repressed

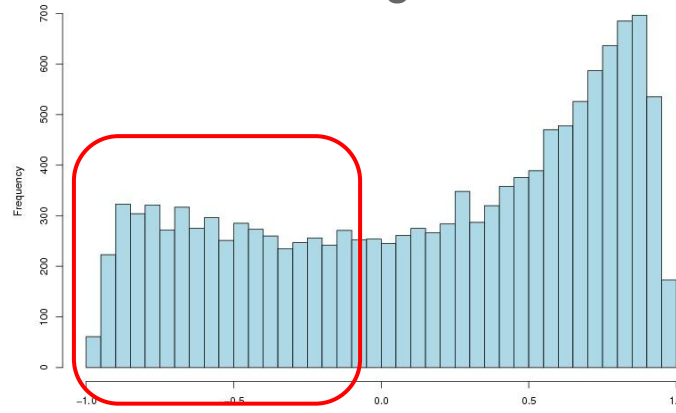
Global consistency between expression, chromatin accessibility, and chromatin conformation!

Exploring correlation between promoter accessibility and gene expression

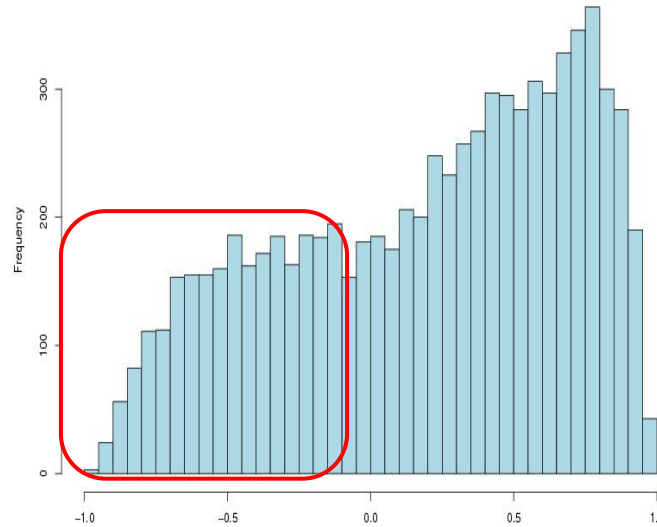
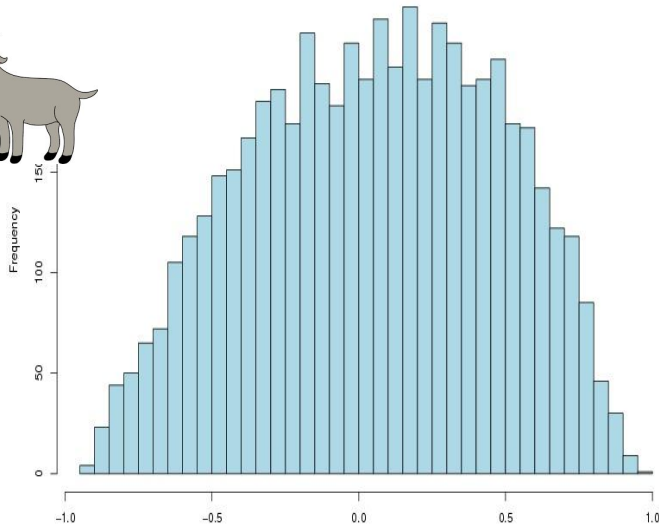
Non-DE genes



DE genes



Positive AND
negative
regulatory
mechanisms?



- Pattern unique to **proximal promoter region** (TSS +/- 1kb)
- Promoter peaks negatively correlated with gene expression tend to be **wider**

Next steps & future work

- **More integrative analyses:**
 - Enhancer / gene relationships
 - Small RNAs vs long RNAs / open chromatin / HiC TADs
- **More comparative analyses:**
 - Evolution of functional elements, in particular regulatory
 - Human/mouse element projection to livestock species
- bioRxiv preprint to be submitted in coming weeks
- More tissues, functional validation and link to genotype/phenotype (G/P) data:
 - **H2020 project proposal** (co-coordinated by E. Giuffra & H. Acloque)
 - . . .

- **Management:**

- Elisabetta Giuffra
 - Sylvain Foissac
 - Sandrine Lagarrigue
 - Marie-Hélène Pinard
- } Coordinators

- **Sampling and assays:**

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- Fany Blanc
- Françoise Drouet
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- Ignacio Gonzalez
- Christophe Klopp
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- Maria Marti
- Kylie Munyard
- Kévin Muret
- David Robelin
- Magali San Cristobal
- Nathalie Villa-Vialaneix
- Matthias Zytnicki

Thanks for your attention!